

QIAquick[®] Nucleotide Removal Kit

The QIAquick Nucleotide Removal Kit (cat. nos. 28304 and 28306) can be stored at room temperature (15–25°C) for up to 12 months if not otherwise stated on label.

Further information

- *QIAquick Spin Handbook*: www.qiagen.com/HB-1196
- Safety Data Sheets: www.qiagen.com/safety
- Technical assistance: support.qiagen.com

Notes before starting

- This protocol is for cleanup of up to 10 µg oligonucleotides (17–40mers) and/or DNA fragments (40 bp to 10 kb) from enzymatic reactions.
- Add isopropanol to Buffer PNI before use (see bottle label for volume).
- Add ethanol (96–100%) to Buffer PE concentrate before use (see bottle label for volume).
- All centrifugation steps are carried out in a conventional tabletop microcentrifuge at room temperature (15–25°C).

1. Add 10 volumes of Buffer PNI to 1 volume of sample and mix. For DNA fragments ≥100 bp, only 5 volumes of Buffer PNI are required.
2. Place a QIAquick spin column in a provided 2 ml collection tube.
3. Apply the sample to the QIAquick spin column and centrifuge for 1 min at 6000 rpm.
4. **For radioactive samples:** Discard the collection tube containing radioactive flow-through appropriately and place the QIAquick spin column into a clean 2 ml collection tube.

For non-radioactive samples: Discard the flow-through and place the QIAquick spin column back into the same collection tube.

5. **For radioactive samples:** Add 500 μ l Buffer PE and centrifuge for 1 min at 3800 \times g (6000 rpm). Discard the flow-through appropriately and repeat the wash with another 500 μ l Buffer PE.
For non-radioactive samples: Add 750 μ l Buffer PE and centrifuge for 1 min at 3800 \times g (6000 rpm).
6. Discard the flow-through and place the QIAquick spin column back in the same collection tube. Centrifuge for an additional 1 min at 17,900 \times g (13,000 rpm). (Residual ethanol from Buffer PE will not be completely removed unless the flow-through is discarded before this additional centrifugation.)
7. Place the QIAquick spin column in a clean 1.5 ml microcentrifuge tube.
8. To elute DNA, add 100–200 μ l Buffer EB (10 mM Tris-Cl, pH 8.5) or water to the center of the QIAquick membrane and centrifuge for 1 min at 17,900 \times g (13,000 rpm). Alternatively, for increased DNA concentration, add 30–50 μ l elution buffer to the center of the QIAquick membrane, let the column stand for 1 min and then centrifuge.
9. If the purified DNA is to be analyzed on a gel, add 1 volume of Loading Dye to 5 volumes of purified DNA. Mix the solution by pipetting up and down before loading the gel.



Scan QR code for handbook.

For up-to-date licensing information and product-specific disclaimers, see the respective QIAGEN kit handbook or user manual.

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