November 2020

Quick-Start Protocol

Procedure for Elution of DNA Extracts from FTA® Elute

Extracted DNA is routinely stored frozen for sample archiving. QIAcard® FTA Elute can be used to stabilize and protect nucleic acids for long-term storage at room temperature (15–25°C). This procedure describes how to apply, store, and elute extracted DNA on FTA Elute cards.

Precautions

Handling: Always wear gloves to avoid contamination of FTA Elute Cards. Follow universal precautions when handling biological specimens.

Storage: Samples can be stored in a multi-barrier pouch with desiccant for long term storage.

Materials required

- QIAcard FTA Elute Indicating Micro (cat. nos. WB120411 and WB120412)
- TE^{-4} buffer (10 mM Tris·Cl, 0.1 mM EDTA)
- Investigator Lyse&Spin Basket Kit (cat. no. 19597 or 19598)
- UniCore Punches 3.00 mm (cat. no. WB100078 or WB100039)
- Cutting Mat 6.0" x 8.0" or 2.5" x 3.0" (cat no. WB100020 or WB100088)
- Heated mixer/shaker
- 1.5 ml microcentrifuge tubes

Procedure

Applying DNA extract to QIAcard FTA Elute Indicating Micro

1. Label the QIAcard FTA Elute Indicating Micro with the appropriate sample identification.
2. Pipette up to 75 µl of DNA extract onto the card within the printed circle area.
3. Allow the sample to air-dry at least 3 h at room temperature until dry.
4. Once completely dried, store the samples in a cool and dry environment until ready to use.

Elution of DNA from QIAcard FTA Elute Indicating Micro

1. Place the FTA Elute Card on a cutting mat.
2. Remove four, 3 mm punches from the FTA Elute Card and place the punches into a single 1.5 ml microcentrifuge tube.

   Note: One to four punches can be used based on known sample concentration and DNA input required for amplification.
3. Pipette 500 µl of TE^{-4} buffer into the microcentrifuge tube containing the 3 mm punches.
4. Close the tube and vortex the microcentrifuge tube for 5 s.
5. Pipette off excess TE^{-4} buffer and discard.
6. Repeat steps 3–5 (for a total of three washes with TE^{-4} buffer).
7. Pipette an appropriate amount of TE^{-4} buffer into the microcentrifuge tube containing the sample punches based on the number of punches and suggested volumes in Table 1.

8. Place the microcentrifuge tube on a heated mixer/shaker at 95°C for 30 min at 1000 rpm.

9. After incubation, briefly centrifuge the microcentrifuge tube to remove any excess liquid from the cap.

10. Transfer the punches and eluate to an Investigator Lyse&Spin Basket and spin at maximum speed for 2 min.

    Note: If there is still liquid in the basket, use a clean pipette tip to move the punches away from the pores of the basket and repeat centrifugation.

11. Remove the basket, discard the punches, and proceed with quantification and/or amplification.

    Note: If the sample is too dilute to meet the DNA input needed for PCR amplification, the sample can be concentrated.

12. Store extracts according to your laboratory protocols.

Table 1. Recommended elution volume (TE^{-4} buffer) for different numbers of card punches

Elution volume	Minimum no. of punches
≥50 µl	1
75 µl	2
100 µl	3
125–150 µl (sample concentration req)	4

Note: Concentration may be required for any combination of punches and elution volumes dependent on the starting concentration of the sample.

Note: Increase the elution volume by 25 µl for every additional 3 mm punch.

Ordering information

Product	Pack size	Cat. no.
QIAcard FTA Elute Indicating Micro	100	WB120411
QIAcard FTA Elute Indicating Micro	25	WB120412
UniCore Punch Kit 3.0 mm	4 (including 2 cutting mats)	WB100039
UniCore Punch 3.0 mm	25	WB100078
Cutting Mat 2.5" × 3.0"	1	WB100088
Cutting Mat 6.0" × 8.0"	1	WB100020
Multi-Barrier Pouches, 3.75″ x 3″	100	WB100036
Indicating Desiccant Pack (1000)	1000 (1 g each)	WB100003
Investigator Lyse&Spin Basket	50	19597
Investigator Lyse&Spin Basket	250	19598

Document Revision History

Date	Changes
11/2020	Initial release