Quick-Start Protocol June 2019

QIAprep® 96 Turbo Miniprep Kit

The QIAprep 96 Turbo Miniprep Kit (cat. nos. 27191 and 27193) can be stored at room temperature (15–25°C) for up to 12 months if not otherwise stated on label.

Further information

- *QIAprep Miniprep Handbook*: **www.qiagen.com/HB-1206**
- Safety Data Sheets: **www.qiagen.com/safety**
- Technical assistance: **support.qiagen.com**

Notes before starting

- Add RNase A solution to Buffer P1, mix, and store at 2–8°C.
- Add ethanol (96–100%) to Buffer PE (see bottle label for volume).
- Do not use 96-well plates that have been damaged.
- Always ventilate the QIAvac 96 slowly after vacuum is switched off.

1. Pellet harvested cells by centrifugation. For growth of bacterial cultures in tubes: >6800 x *g* for 3 min; for growth in blocks: 2100 x *g* for 5 min.
2. Resuspend pelleted bacterial cells in 250 µl Buffer P1 and transfer to the provided S-Block (if cells were not harvested in this block).
3. Add 250 µl Buffer P2 to each sample. Dry the top of the S-Block with a paper towel, seal the block with the tape provided, gently invert the block 4–6 times to mix, and incubate at room temperature for 5 min.
4. During incubation, place the TurboFilter® 96 plate in the QIAvac 96 top plate. Place the plate holder inside the QIAvac base. Place the QIAprep 96 plate into the plate holder. Place the QIAvac top plate squarely over the base.
5. After incubation, remove tape, add 350 µl Buffer N3, dry the top of the block with a paper towel and seal the block with new tape. Gently invert the block 4–6 times.

6. Remove the tape and pipet the lysates from step 5 into the wells of the TurboFilter plate. Apply vacuum until all samples have passed through.
7. Switch off vacuum. Discard the TurboFilter plate. Transfer the QIAprep plate to the top plate of the manifold. Replace the plate holder in the base with a waste tray. Place the top plate squarely over the base. Apply vacuum.
8. **Recommended**: Switch off vacuum and wash the QIAprep plate by adding 0.9 ml Buffer PB to each well. Apply vacuum.
9. Switch off vacuum. Wash the QIAprep plate by adding 0.9 ml of Buffer PE to each well and apply vacuum. Repeat once.
10. After Buffer PE has been drawn through all wells, apply maximum vacuum for an additional 10 min to dry the membrane.
11. Switch off vacuum. Lift the top plate from the base (not the QIAprep plate from the top plate). Vigorously tap the top plate on a stack of absorbent paper until no drops come out, and then blot the nozzles of the QIAprep plate with clean absorbent paper. Proceed with either step 12.a or 12.b.
12. **a) For elution into provided collection tubes:** Replace the waste tray with the blue collection microtube rack. Place the top plate back on the base.

    b) For elution into a 96-well microplate: Replace waste tray with an empty blue collection microtube rack. Place 96-well microplate directly on rack. Place top plate back on base.
13. To elute DNA, add 100 µl of Buffer EB or water to the center of each well of the QIAprep plate, let it stand for 1 min, and apply maximum of vacuum for 5 min. Switch off vacuum.

Revision History

Document	Changes	Date
HB-0610-003	Replaced flat-bottom blocks with S-Blocks.	June 2019

Scan QR code for handbook.

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